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CYLINDOL A, A NOVEL BIPHENYL ETHER WITH 5-LIPOXYGENASE INHIBITORY ACTIVITY, AND A RELATED COMPOUND FROM *IMPERATA CYLINDRICA*

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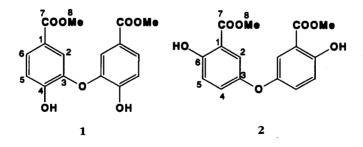
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ABSTRACT.—Cylindol A [1] and B [2], two novel substances, have been isolated from *Imperata cylindrica*, and their structures have been elucidated on the basis of their spectral data coupled with chemical evidence and total synthesis. Cylindol A [1] showed 5-lipoxygenase inhibitory activity.

In the course of our investigations of pharmacologically active substances from medicinal plants, we have devoted our attention to the occurrence of natural products having 5-lipoxygenase inhibitory activity since these compounds may be useful as anti-inflammatory drugs in therapeutic applications.

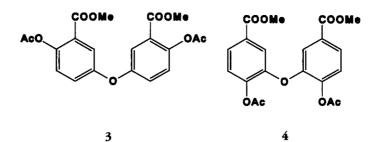
The rhizomes of *Imperata cylindrica* Beauvois (Gramineae) (Japanese name "Chigaya") have been used in Chinese medicine as diuretic and anti-inflammatory agents (1-3). However, only a few studies concerning the constituents of this plant have been conducted and the presence of migrated hopane and arborane triterpenoids was reported by Nishimoto *et al.* (4–7). In this paper, we describe the isolation of the new biphenyl ether compounds, cylindols A [1] and B [2], from

An EtOAc-soluble extract of the rhizomes of I. cylindrica was repeatedly chromatographed over Si gel to afford cylindols A [1] and B [2]. Cylindol A [1] showed a molecular ion at m/z 318 in the eims. The ir spectrum of **1** displayed absorption bands at 3360 and 1695 $\rm cm^{-1}$, indicating the presence of OH and ester functionalities in the molecule. The presence of a 1,2,4-tri-substituted phenyl ring was elucidated based on the ¹³C-nmr data and the coupling pattern of aromatic protons in the ¹H-nmr spectrum. From an analysis of the eims and ¹³C-nmr spectra, the structure was indicated as a symmetrical biphenyl ether. Cylindol B [2] was considered to be an isomer of cylindol A [1] from its ir and nmr spectra. In the ms spectrum, however, a molecular ion typical of a dimeric structure was not



I. cylindrica, and the total synthesis and 5lipoxygenase inhibitory activity of cylindol A [**1**]. detected but only a monomer ion was observed at m/z 168. When cylindol B [1] was acetylated by treatment with Ac₂O and pyridine, acetylcylindol B [3] showed a molecular ion at m/z 402. The chemical shifts and splitting patterns in

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the ¹H-nmr spectrum of acetylcylindol B [3] pointed to the presence of a 1,2,4-trisubstituted phenyl ring. Therefore, cylindol B [2] was considered to be a symmetrical 1,2,4-tri-substituted biphenyl ether compound like cylindol A [1]. Cylindol A [1] was also acetylated, and acetylcylindol A [4] showed a molecular ion at m/z 402. In these two acetates, the acetylation shifts (ca. 0.35 ppm at H-5 in cylindol A, 0.28 ppm in cylindol B) indicated two possible relative locations of the functionalities in each molecule. The tentative structures 1 and 2 were proposed for cylindol A and B. However, it was not unambiguously determined which structure corresponded to which compound. Therefore, the structure of 1 was confirmed by total synthesis. As shown in Figure 1, compounds 5 and 6 were coupled to give a biphenyl ether, and then oxidation, demethylation, and esterification reactions were performed to yield the target compound. The spectral data of the synthetic compound were completely identical with those of cylindol A [1]. In this manner, the structures of cylindol A and B were determined as 1and 2, respectively.

5-Lipoxygenase was inhibited by cylindol A [1] (64% at 30 μ M; known standard, phenidone, 50% at 15 μ M), but not by cylindol B [2]. It is possible, therefore, that cylindol A [1] is one of the anti-inflammatory principles of *I. cylindrica*. Further detailed studies on the biological activity are now underway.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES .--- The

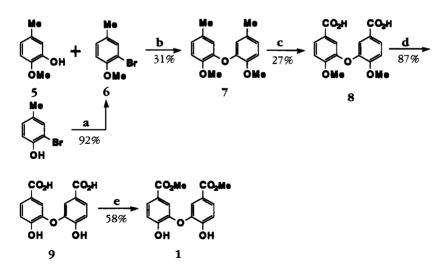


FIGURE 1. Total Synthesis of Cylindol A [1]. a) Me₂SO₄/NaOH/EtOH/110°. b) K₂CO₃/ Kl/Cu/DMSO/160°. c) KMnO₄/KOH/H₂O/reflux. d) BBr₃/CH₂Cl₂/0°~room temperature. e) concentrated H₂SO₄/MeOH/reflux.

it spectra were taken on a Shimadzu IR-408 spectrometer. Mps were determined on a Yamaco micro-melting point apparatus and are uncorrected. Uv spectra were taken on a Hitachi U-2000 spectrometer. Eims were obtained on a JEOL JMS DX-303 or a JEOL JMS D-100 mass spectrometer. ¹H- and ¹³C-nmr spectra were recorded on JEOL JNM GX-500, Hitachi R-3000, and JEOL JNM EX-270 nmr spectrometers using TMS as an internal standard. ¹³C-Nmr multiplicities were determined by off-resonance decoupling experiments. Synthetic starting materials were supplied by Tokyo Kasei Co., Ltd.

PLANT MATERIAL.—The rhizomes of *l. cylindrica* were supplied by Nippon Hunmatsu Yakuhin, Ltd., Osaka, Japan. A voucher specimen has been deposited in the herbarium of the Pharmaceutical Institute of Tohoku University.

EXTRACTION AND ISOLATION.—The rhizomes of *I. cylindrica* (2 kg) were cut and extracted with 5 liters of boiling MeOH and 5 liters of boiling H₂O. The residues from the MeOH and H₂O extracts (280 g and 70 g) were combined, then partitioned between EtOAc and H₂O. The EtOAc extract was concentrated *in vacuo*.

The EtOAc extract (35 g) was directly chromatographed on Si gel. After elution of higher fatty acids and their esters with CHCl₃-MeOH (100:1), a pale yellow oil (110 mg) was eluted by a solution of CHCl₃-MeOH (20:1), which was repeatedly chromatographed by Si gel prep. tlc using solutions of CHCl₃-MeOH (15:1) and EtOAc-MeOH(60:1) as eluates, to afford cylindols A [1] (7 mg) and B [2] (5 mg), in 0.00035% and 0.00025% yield, respectively.

Cylindol A [1] was isolated as a colorless amorphous powder; mp 217–219°; ir (film) ν max 3360, 1695 cm⁻¹; uv (MeOH) λ max (log ϵ) 220 (4.55), 257 (4.39), 288 (4.02) nm; ¹H nmr (CD₃OD, 500 MHz) δ 3.90 (6H, s, 2×CH₃-8), 7.09 (2H, d, J=7.9 Hz, 2×H-5), 7.52 (2H, d, J=1.8 Hz, 2×H-2), 7.80 (2H, dd, J=7.9 and 1.8 Hz, 2×H-6); ¹³C nmr (CD₃OD, 125 MHz) δ 52.5 (q, C-8), 118 (d, C-5), 121.5 (d, C-2), 123 (s, C-1), 128 (d, C-6), 145.5 (s, C-3), 154.8 (s, C-4), 168 (s, C-7), eims m/z [M]⁺ 318 (60), 286 (100), 269 (45), 255 (37), 128 (30); hreims: found 318.0742 (calcd for C₁₆H₁₄O₇ 318.0740).

Cylindol B [2] was isolated as a colorless amorphous powder; mp 187–190°; ir (film) ν max 3540, 1695 cm⁻¹; uv (MeOH) λ max (log ϵ) 220 (4.55), 257 (4.39), 288 (4.02) nm; ¹H nmr(CD₃OD, 300 MHz) δ 3.98 (6H, s, 2×CH₃-8), 6.93 (2H, d, J=9.6 Hz, 2×H-5), 7.64 (4H, complex, 2×H-2, 2×H-4); ¹³C nmr (CD₃OD, 75 MHz) δ 56.4 (q, C-8), 114 (d, C-5), 116 (d, C-2), 123 (s, C-1), 125 (d, C-4), 149 (s, C-3), 153 (s, C-6), 170 (s, C-7).

Acetylation of cylindol A.—To a solution of 1 mg of cylindol A [1] in 1 ml of pyridine, 0.5 ml of Ac₂O was added at room temperature. After 3 h, the reaction mixture was azeotroped with toluene to give acetylcylindol A [4]; ir (film) ν max 1770, 1725 cm⁻¹; eims *m*/*z* [M]⁺ 402 (45), 318 (30), 168 (100); ¹H nmr (CD₃OD, 500 MHz) δ 2.32 (6H, s, 2×OAc), 3.96 (6H, s, 2×CH₃-8), 7.44 (2H, d, *J*=7.9 Hz, 2×H-5), 7.65 (2H, d, *J*=1.8 Hz, 2×H-2), 7.97 (2H, dd, *J*=7.9 and 1.8 Hz, 2×H-6).

Acetylation of cylindol B.—To a solution of 1 mg of cylindol B [2] in 0.5 ml of pyridine, 0.5 ml of Ac₂O was added at room temperature. After stirring 3 h, the reaction mixture was azeotroped with toluene and purified by Si gel tlc using CHCl₃-MeOH (30:1) as eluate: ir (film) ν max 1765, 1685 cm⁻¹; eims m/z [M]⁺ 402 (10), 318 (30), 168 (100), 153 (35); ¹H nmr (CD₃OD, 300 MHz) δ 2.36(6H, s2×OAc), 3.95(6H, s, 2×CH₃-8), 7.21 (2H, dJ=8.1 Hz, 2×H-5), 7.72 (2H, dd, J=8.1 and 2.3 Hz, 2×H-4), 7.78 (2H, d, J=2.3 Hz, 2×H-2).

Methylation of 2-bromo-p-cresol.—A solution of 25.8 g (138 mmol) of 2-bromo-p-cresol in 50 ml of EtOH and 21.1 g (167 mmol) of Me_2SO_4 was treated with aqueous NaOH (8.6 g in 15 ml of H_2O) at 0°. The reaction mixture was heated to 110°. After 4 h, the reaction mixture was concentrated *in vacuo* and partitioned between Et₂O and 5% HCl. The organic layer was washed with brine, dried over MgSO₄, and concentrated *in vacuo* to give 25.4 g (92%) of **6**; ¹H nmr (CDCl₃, 270 MHz) δ 1.56 (3H, s), 3.85 (3H, s), 6.78 (1H, d, J=8.3 Hz), 7.05 (1H, dd, J=8.3 and 2.0 Hz), 7.35 (1H, d, J=2.0 Hz); ¹³C nmr (CDCl₃, 67.8 MHz) δ 20.1, 56.3, 111.3, 111.8, 128.9, 131.4, 133.7, 153.7.

Coupling reaction of 2-methoxy-5-methyl phenol [5] and 6.—To a solution of 5 g (36.2 mmol) of 2methoxy-5-methyl phenol [5] and 8.73 g (43.4 mmol) of 6 in 10 ml of DMSO were added 7.5 g (54.3 mmol) of K_2CO_3 , 1.2 g (7.2 mmol) of KI, and 280 mg of Cu powder. The reaction mixture was heated to 150°. After 3 h, the reaction mixture was diluted with Et₂O, washed with H₂O, 5% HCl, and brine. The organic layer was dried over MgSO₄, concentrated *in vacuo* and chromatographed on Si gel (C₆H₁₄-EtOAc, 10:1) to give 2.9 g (31%) of 7; ¹H nmr (CDCl₃, 270 MHz) δ 2.49 (6H, s), 3.82 (6H, s), 6.63 (2H, br s), 6.85 (4H, br s); ¹³C nmr (CDCl₃, 67.8 MHz) δ 20.7, 56.2, 112.6, 119.6, 123.9, 130.6, 145.8, 148.4.

Oxidation of 7.—To a suspension of 2.2 g (8.55 mmol) of 7 in 200 ml of H₂O were added 13.5 g (85.5 mmol) of KMnO₄ and 7.18 g (128 mmol) of KOH. The reaction mixture was refluxed for 10 h, filtered, cooled to 0°, and acidified with concentrated HCl to give 745 mg (27%) of **8**; ¹H nmr (DMSO-d₆, 270 MHz) δ 3.96 (6H, s), 7.35 (2H, d, J=8.6 Hz), 7.36 (2H, d, J=2.0 Hz), 7.86 (2H, dd, J=8.6 and 2.0 Hz), 12.9 (2H, br s); ¹³C nmr (DMSO-*d*₆, 67.8 MHz) **§** 55.9, 112.7, 118.7, 123.0, 126.4, 144.4, 153.8, 166.3.

Demetbylation of 8.—To a stirred suspension of 400 mg (1.26 mmol) of 8 in 3 ml of CH_2Cl_2 was added 0.6 ml (6.3 mmol) of BBr₃ at 0°. After 3 h, the reaction mixture was poured into ice-cooled H₂O and the precipitate was filtered to give 316 mg (86%) of 9; ¹H nmr (DMSO-d₆, 270 MHz) δ 7.11 (2H, d, J=8.3 Hz), 7.35 (2H, d, J=2.0 Hz), 7.70 (2H, dd, J=8.3 and 2.0 Hz), 10.5 (2H, br s).

Esterification of 9.—To a solution of 300 mg (1.03 mmol) of 9 in 30 ml of anhydrous MeOH was added 0.1 ml of concentrated H₂SO₄. The reaction mixture was refluxed for 5 h, concentrated *in vacuo*, and partitioned between Et₂O and H₂O. An organic layer was washed with brine, dried over MgSO₄, and concentrated *in vacuo*. Purification by Si gel cc (CHCl₃-MeOH, 50:1) afforded 192 mg (58%) of cylindol A [1] as an amorphous powder; mmp with the natural compound, 217–219°.

5-Lipoxygenase inhibitory activity of cylindol A [1].—The 5-lipoxygenase assay was run using a crude enzyme preparation from rat basophilic leukemia cells (RBL-1). Cylindol A was pre-incubated with the enzyme for 5 min at room temperature and the reaction was initiated by addition of arachidonic acid. Following 8 min incubation at room temperature, the reaction was terminated by addition of 2 ml of 1 N NaOH, and absorbance read at 234 nm to determine levels of 5-HETE (8,9).

LITERATURE CITED

- 1. K. Yamaguchi, Chosen Igakukaishi, 85, 173 (1928).
- 2. T. Haginiwa and M. Harada, Shoyakugaku Zasshi, 17, 6 (1963).
- 3. P. Kancharapee, Shoyakugaku Zasshi, 21, 65 (1967).
- T. Ohmoto, K. Nishimoto, M. Ito, and S. Natori, Chem. Pharm. Bull., 13, 224 (1965).
- 5. K. Nishimoto, M. Ito, S. Natori, and T. Ohmoto, *Tetrabedron Lett.*, 2245 (1965).
- K. Nishimoto, M. Ito, S. Natori, and T. Ohmoto, Chem. Pharm. Bull., 14, 97 (1966).
- K. Nishimoto, M. Ito, S. Natori, and T. Ohmoto, *Tetrabedron*, 24, 735 (1968).
- T. Shimuzu, O. Radmark, and B. Samuelsson, Proc. Natl. Acad. Sci. U.S.A., 81, 689 (1984).
- R.W. Egan and P.H. Gale, J. Biol. Chem., 260, 11554 (1985).

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